

*Sample size by simulation for
clinical trials with survival outcomes:
the **simsam** package in action*

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The **simsam** package

simsam uses simulation to determine the sample size required to achieve given statistical power to detect a given effect, for *any* hypothesis test under *any* statistical model that can be programmed in Stata.

**Hooper R. Versatile sample size calculation using simulation.
Stata Journal 2013;13(1):21-38**



Why worry about sample size?

“The number of subjects in a clinical trial should always be large enough to provide a reliable answer to the questions addressed. This number is usually determined by the primary objective of the trial.”

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“This [sample size calculation] is frequently one of the least credible components of a trial [funding] application.”

UK National Institute for Health Research



Basic syntax of **simsam**

```
. simsam subcommand_name n_name, ///  
> detect (parameter_name (parameter_value)) ///  
> null (parameter_name (null_value)) ///  
> assuming (nuisance_parameter1 (par1_value) ... ) ///  
> p(.8) inc(10) prec(0.01)
```

where *subcommand_name* is the name of a user-written program which codes the statistical model and the hypothesis test



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NB **simsam** doesn't do anything by itself – **it needs software**



A modular view of a **simsam** subcommand

```
program define subcommand_name, rclass
```

```
  syntax , n_name(integer)  ///  
    parameter_name(real)  ///  
    nuisance_parameter1(real)  ///  
  :
```

```
  drop _all
```

```
  [generate data-set]
```

```
  [analyse data-set]
```

```
  return scalar p = expression_for_pvalue
```

```
end
```

Something more complex: a two-stage adaptive design

```
program define subcommand_name, rclass
```

```
  syntax , ...
```

```
  drop _all
```

```
[generate data from stage 1]
```

```
[analyse data from stage 1 and calculate p-value]
```

```
[choose to stop there, or else adapt the protocol based  
on stage 1 results, then generate data from stage 2]
```

```
[analyse data from stage 2 and calculate p-value]
```

```
[return a combined p-value from the two stages]
```

```
end
```

Trials with survival (time-to-event) outcomes

For an individually-randomised trial where the outcome is time until death (possibly censored), the total number of deaths that must be observed to detect hazard ratio Δ with given power is approximately (Schoenfeld, 1983)

$$4(z_{\beta} + z_{1-\alpha/2}) / \log^2 \Delta$$

Jahn-Eimermacher et al (2011) extend this to cluster-randomised trials analysed with frailty models, for which the above formula underestimates sample size. Their extended formula still underestimates sample size when the cluster size is variable.



```
program define s_survival, rclass
```

```
    syntax , recrdur(integer) recrrate(integer) ///  
           hr(real) failratec(real) ///  
           folldur(real) droprate(real)
```

```
    drop _all
```

```
    set obs `='recrdur'*`recrrate''
```

```
    gen group=mod(_n,2)
```

```
    gen abs_trecr=sum(-log(runiform()))/`recrrate'  
    gen tfail=-log(runiform())/`failratec'*`hr'^group  
    gen tdrop=-log(runiform())/`droprate'  
    gen tstop=`recrdur'+`folldur'-abs_trecr  
    drop if tstop<0
```

```
    gen t=min(tfail, tdrop, tstop)  
    gen fail=(t<min(tdrop, tstop))  
    stset t, failure(fail)
```

```
    stcox group
```

```
    return scalar p=2*normal(-abs(_b[group]/_se[group]))
```

```
end
```

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- Simplest approach: treat the result as non-significant.
 - To do this you just need to exit the subcommand without returning a p-value
 - A general approach is to encase the analysis "module" in **capture noisily** brackets



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```
capture noisily {  
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}
```



Capturing errors in the survival analysis

- Assuming the data are legitimate, the only error you are really likely to encounter with `stcox` is failure to converge.
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Capturing errors in the survival analysis

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- **You have to decide how you would handle errors if they occurred in the analysis of the real data.**
- *i.e.* you need to specify an Analysis Plan



Capturing errors in the survival analysis

e.g. if Cox regression fails to converge, try parametric regression with a Weibull model for survival times

```
capture noisily {
    stcox group
    return scalar p=2*normal(-abs(_b[group]/_se[group]))
}
if _rc~=0 {
    streg group, dist(weibull)
    return scalar p=2*normal(-abs(_b[group]/_se[group]))
}
```



Convergence problems that don't lead to errors: controlling the number of iterations used for estimation

- Generally **stcox** converges after a few iterations
- Very occasionally it will continue on to the maximum number of iterations (16,000 by default) without producing a non-convergence error
- Hence **simsam** will appear to be hung up but will not halt with an error message



Convergence problems that don't lead to errors: controlling the number of iterations used for estimation

The solution is to re-set the maximum number of iterations:

```
. set maxiter 20
. simsam s_survival recrrate, ///
>     detect(hr(1.5)) null(hr(1.0))   ///
>     assuming(failratec(0.5) ///
>             recrdur(2) folldur(1))   ///
>     p(.8) inc(1) prec(0.001)
```



iteration	recrrate	power	(99% CI)
1	100	0.6500	(0.5172, 0.7681)
2	143	0.8120	(0.7782, 0.8428)
3	139	0.7971	(0.7866, 0.8074)
4	141	0.8004	(0.7972, 0.8037)
5	141	0.8009	(0.7999, 0.8019)
6	140	0.7988	(0.7978, 0.7998)
null	141	0.0499	(0.0489, 0.0509)

recrrate = 141
 achieves 80.09% power (99% CI 79.99, 80.19)
 at the 5% significance level
 to detect
 hr = 1.5
 assuming
 failratec = 0.5
 recrdur = 2
 folldur = 1

 under null: 4.99% power (99% CI 4.89, 5.09)

Concluding remarks

Simulation for sample size calculation

- is accurate and versatile
- but must anticipate every contingency
- needs statistician input
- forces you to think about the analysis in detail (no bad thing)
- helps others to develop related applications



- More info at <http://webpace.qmul.ac.uk/rrhooper/simsam>
- simsam update planned for Jan 2014

Thank you

