

**clogit** — Conditional (fixed-effects) logistic regression[Description](#)  
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## Description

`clogit` fits a conditional logistic regression model for matched case–control data, also known as a fixed-effects logit model for panel data. `clogit` can compute robust and cluster–robust standard errors and adjust results for complex survey designs.

See [\[R\] `asclogit`](#) if you want to fit McFadden’s choice model (McFadden 1974).

## Quick start

Conditional logistic regression model of `y` on `x` with matched case–control pairs data identified by `idvar`

```
clogit y x, group(idvar)
```

Fixed-effects logistic regression model with panels identified by `idvar`

```
clogit y x, group(idvar)
```

Add categorical variable `a` and report results as odds ratios

```
clogit y x i.a, or
```

As above, but using sampling probability weight `wvar`

```
clogit y x i.a [pweight = wvar], or
```

## Menu

Statistics > Categorical outcomes > Conditional logistic regression

## Syntax

```
clogit depvar [indepvars] [if] [in] [weight], group(varname) [options]
```

*depvar* is treated as binary regardless of values; *depvar* equal to nonzero and nonmissing (typically equal to 1) indicates a positive outcome, whereas *depvar* equal to 0 indicates a negative outcome.

<i>options</i>	Description
Model	
* <u>group</u> ( <i>varname</i> )	matched group variable
<u>offset</u> ( <i>varname</i> )	include <i>varname</i> in model with coefficient constrained to 1
<u>constraints</u> ( <i>constraints</i> )	apply specified linear constraints
<u>collinear</u>	keep collinear variables
SE/Robust	
<u>vce</u> ( <i>vcetype</i> )	<i>vcetype</i> may be <code>oim</code> , <code>robust</code> , <code>cluster</code> <i>clustvar</i> , <code>opg</code> , <code>bootstrap</code> , or <code>jackknife</code>
<code>nonest</code>	do not check that panels are nested within clusters
Reporting	
<u>level</u> (#)	set confidence level; default is <code>level(95)</code>
<code>or</code>	report odds ratios
<u>nocnsreport</u>	do not display constraints
<u>display_options</u>	control columns and column formats, row spacing, line width, display of omitted variables and base and empty cells, and factor-variable labeling
Maximization	
<u>maximize_options</u>	control the maximization process; seldom used
<u>coeflegend</u>	display legend instead of statistics

\*group(*varname*) is required.

*indepvars* may contain factor variables; see [U] 11.4.3 Factor variables.

`bayes`, `bootstrap`, `by`, `fp`, `jackknife`, `mfp`, `mi estimate`, `nestreg`, `rolling`, `statsby`, `stepwise`, and `svy` are allowed; see [U] 11.1.10 Prefix commands. For more details, see [BAYES] bayes: clogit.

`vce(bootstrap)` and `vce(jackknife)` are not allowed with the `mi estimate` prefix; see [MI] mi estimate.

Weights are not allowed with the `bootstrap` prefix; see [R] bootstrap.

`vce()`, `nonest`, and weights are not allowed with the `svy` prefix; see [SVY] svy.

`fweights`, `iweights`, and `pweights` are allowed (see [U] 11.1.6 weight), but they are interpreted to apply to groups as a whole, not to individual observations. See Use of weights below.

`coeflegend` does not appear in the dialog box.

See [U] 20 Estimation and postestimation commands for more capabilities of estimation commands.

## Options

Model

group(*varname*) is required; it specifies an identifier variable (numeric or string) for the matched groups. strata(*varname*) is a synonym for group(*varname*).

offset(*varname*), constraints(*constraints*), collinear; see [R] estimation options.

SE/Robust

`vce(vcetype)` specifies the type of standard error reported, which includes types that are derived from asymptotic theory (`oim`, `opg`), that are robust to some kinds of misspecification (`robust`), that allow for intragroup correlation (`cluster clustvar`), and that use bootstrap or jackknife methods (`bootstrap`, `jackknife`); see [R] [vce\\_option](#).

`nonest`, available only with `vce(cluster clustvar)`, prevents checking that matched groups are nested within clusters. It is the user's responsibility to verify that the standard errors are theoretically correct.

Reporting

`level(#)`; see [R] [estimation options](#).

`or` reports the estimated coefficients transformed to odds ratios, that is,  $e^b$  rather than  $b$ . Standard errors and confidence intervals are similarly transformed. This option affects how results are displayed, not how they are estimated. `or` may be specified at estimation or when replaying previously estimated results.

`nocnsreport`; see [R] [estimation options](#).

`display_options`: `nocl`, `nopvalues`, `noomitted`, `vsquish`, `noemptycells`, `baselevels`, `allbaselevels`, `nofvlabel`, `fvwrap(#)`, `fvwrapon(style)`, `cformat(%fmt)`, `pformat(%fmt)`, `sformat(%fmt)`, and `nolstretch`; see [R] [estimation options](#).

Maximization

`maximize_options`: `difficult`, `technique(algorithm_spec)`, `iterate(#)`, `[no]log`, `trace`, `gradient`, `showstep`, `hessian`, `showtolerance`, `tolerance(#)`, `ltolerance(#)`, `nrtolerance(#)`, `nonrtolerance`, and `from(init_specs)`; see [R] [maximize](#). These options are seldom used.

Setting the optimization type to `technique(bhhh)` resets the default `vcetype` to `vce(opg)`.

The following option is available with `clgit` but is not shown in the dialog box:

`coeflegend`; see [R] [estimation options](#).

## Remarks and examples

[stata.com](http://www.stata.com)

Remarks are presented under the following headings:

- [Introduction](#)
- [Matched case-control data](#)
- [Use of weights](#)
- [Fixed-effects logit](#)

## Introduction

`clgit` fits maximum likelihood models with a dichotomous dependent variable coded as 0/1 (more precisely, `clgit` interprets 0 and not 0 to indicate the dichotomy). Conditional logistic analysis differs from regular logistic regression in that the data are grouped and the likelihood is calculated relative to each group; that is, a conditional likelihood is used. See [Methods and formulas](#) at the end of this entry.

Biostatisticians and epidemiologists call these models conditional logistic regression for matched case–control groups (see, for example, [Hosmer, Lemeshow, and Sturdivant \[2013, chap. 7\]](#)) and fit them when analyzing matched case–control studies with 1:1 matching, 1: $k_{2i}$  matching, or  $k_{1i}$ : $k_{2i}$  matching, where  $i$  denotes the  $i$ th matched group for  $i = 1, 2, \dots, n$ , where  $n$  is the total number of groups. `clogit` fits a model appropriate for all of these matching schemes or for any mix of the schemes because the matching  $k_{1i}$ : $k_{2i}$  can vary from group to group. `clogit` always uses the true conditional likelihood, not an approximation. Biostatisticians and epidemiologists sometimes refer to the matched groups as “strata”, but we will stick to the more generic term “group”.

Economists and other social scientists typically call the model fit by `clogit` a fixed-effects logit model for panel data (see, for example, [Chamberlain \[1980\]](#)). The data used to fit a fixed-effects logit model look exactly like the data biostatisticians and epidemiologists call  $k_{1i}$ : $k_{2i}$  matched case–control data. In terms of how the data are arranged,  $k_{1i}$ : $k_{2i}$  matching means that in the  $i$ th group, the dependent variable is 1 a total of  $k_{1i}$  times and 0 a total of  $k_{2i}$  times. There are a total of  $T_i = k_{1i} + k_{2i}$  observations for the  $i$ th group. This data arrangement is what economists and other social scientists call “panel data”, “longitudinal data”, or “cross-sectional time-series data”.

So no matter what terminology you use, the computation and the use of the `clogit` command is the same. The following example shows how your data should be arranged to use `clogit`.

### ► Example 1

Suppose that we have grouped data with the variable `id` containing a unique identifier for each group. Our outcome variable, `y`, contains 0s and 1s. If we were biostatisticians,  $y = 1$  would indicate a case,  $y = 0$  would be a control, and `id` would be an identifier variable that indicates the groups of matched case–control subjects.

If we were economists,  $y = 1$  might indicate that a person was unemployed at any time during a year and  $y = 0$ , that a person was employed all year, and `id` would be an identifier variable for persons.

If we list the first few observations of this dataset, it looks like

```
. use http://www.stata-press.com/data/r15/clogitid
. list y x1 x2 id in 1/11
```

	y	x1	x2	id
1.	0	0	4	1014
2.	0	1	4	1014
3.	0	1	6	1014
4.	1	1	8	1014
5.	0	0	1	1017
6.	0	0	7	1017
7.	1	1	10	1017
8.	0	0	1	1019
9.	0	1	7	1019
10.	1	1	7	1019
11.	1	1	9	1019

Pretending that we are biostatisticians, we describe our data as follows. The first group (`id = 1014`) consists of four matched persons: 1 case ( $y = 1$ ) and three controls ( $y = 0$ ), that is, 1:3 matching. The second group has 1:2 matching, and the third 2:2.

Pretending that we are economists, we describe our data as follows. The first group consists of 4 observations (one per year) for person 1014. This person had a period of unemployment during 1 year of 4. The second person had a period of unemployment during 1 year of 3, and the third had a period of 2 years of 4.

Our independent variables are  $x_1$  and  $x_2$ . To fit the conditional (fixed-effects) logistic model, we type

```
. clogit y x1 x2, group(id)
note: multiple positive outcomes within groups encountered.
Iteration 0:  log likelihood = -123.42828
Iteration 1:  log likelihood = -123.41386
Iteration 2:  log likelihood = -123.41386
Conditional (fixed-effects) logistic regression

                                Number of obs   =          369
                                LR chi2(2)       =           9.07
                                Prob > chi2      =          0.0107
                                Pseudo R2        =          0.0355
Log likelihood = -123.41386
```

y	Coef.	Std. Err.	z	P> z	[95% Conf. Interval]
x1	.653363	.2875215	2.27	0.023	.0898312 1.216895
x2	.0659169	.0449555	1.47	0.143	-.0221943 .1540281



### □ Technical note

The message “note: multiple positive outcomes within groups encountered” at the top of the `clogit` output for the previous example merely informs us that we have  $k_{1i} : k_{2i}$  matching with  $k_{1i} > 1$  for at least one group. If your data should be  $1 : k_{2i}$  matched, this message tells you that there is an error in the data somewhere.

We can see the distribution of  $k_{1i}$  and  $T_i = k_{1i} + k_{2i}$  for the data of the [example 1](#) by using the following steps:

```
. by id, sort: generate k1 = sum(y)
. by id: replace k1 = . if _n < _N
(303 real changes made, 303 to missing)
. by id: generate T = sum(y<.)
. by id: replace T = . if _n < _N
(303 real changes made, 303 to missing)
. tabulate k1
```

k1	Freq.	Percent	Cum.
1	48	72.73	72.73
2	12	18.18	90.91
3	4	6.06	96.97
4	2	3.03	100.00
Total	66	100.00	

```
. tabulate T
```

T	Freq.	Percent	Cum.
2	5	7.58	7.58
3	5	7.58	15.15
4	12	18.18	33.33
5	11	16.67	50.00
6	13	19.70	69.70
7	8	12.12	81.82
8	3	4.55	86.36
9	7	10.61	96.97
10	2	3.03	100.00
Total	66	100.00	

We see that  $k_{1i}$  ranges from 1 to 4 and  $T_i$  ranges from 2 to 10 for these data. □

### □ Technical note

For  $k_{1i} : k_{2i}$  matching (and hence in the general case of fixed-effects logit), `clogit` uses a recursive algorithm to compute the likelihood, which means that there are no limits on the size of  $T_i$ . However, computation time is proportional to  $\sum T_i \min(k_{1i}, k_{2i})$ , so `clogit` will take roughly 10 times longer to fit a model with 10:10 matching than one with 1:10 matching. But `clogit` is fast, so computation time becomes an issue only when  $\min(k_{1i}, k_{2i})$  is around 100 or more. See [Methods and formulas](#) for details. □

## Matched case–control data

Here we give a more detailed example of matched case–control data.

### ▷ Example 2

[Hosmer, Lemeshow, and Sturdivant \(2013, 24\)](#) present data on matched pairs of infants, each pair having one with low birthweight and another with regular birthweight. The data are matched on age of the mother. Several possible maternal exposures are considered: race (three categories), smoking status, presence of hypertension, presence of uterine irritability, previous preterm delivery, and weight at the last menstrual period.

```
. use http://www.stata-press.com/data/r15/lowbirth2, clear
(Applied Logistic Regression, Hosmer & Lemeshow)
. describe
Contains data from http://www.stata-press.com/data/r15/lowbirth2.dta
  obs:           112                Applied Logistic Regression,
                                      Hosmer & Lemeshow
vars:           9                   30 Jan 2016 08:46
size:          1,120
```

variable name	storage type	display format	value label	variable label
pairid	byte	%8.0g		Case-control pair ID
low	byte	%8.0g		Baby has low birthweight
age	byte	%8.0g		Age of mother
lwt	int	%8.0g		Mother's last menstrual weight
smoke	byte	%8.0g		Mother smoked during pregnancy
ptd	byte	%8.0g		Mother had previous preterm baby
ht	byte	%8.0g		Mother has hypertension
ui	byte	%8.0g		Uterine irritability
race	byte	%9.0g	race	race of mother: 1=white, 2=black, 3=other

Sorted by:

We list the case-control indicator variable, low; the match identifier variable, pairid; and two of the covariates, lwt and smoke, for the first 10 observations.

```
. list low lwt smoke pairid in 1/10
```

	low	lwt	smoke	pairid
1.	0	135	0	1
2.	1	101	1	1
3.	0	98	0	2
4.	1	115	0	2
5.	0	95	0	3
6.	1	130	0	3
7.	0	103	0	4
8.	1	130	1	4
9.	0	122	1	5
10.	1	110	1	5

## 8 clogit — Conditional (fixed-effects) logistic regression

We fit a conditional logistic model of low birthweight on mother's weight, race, smoking behavior, and history.

```
. clogit low lwt smoke ptd ht ui i.race, group(pairid) nolog
Conditional (fixed-effects) logistic regression

                                Number of obs   =       112
                                LR chi2(7)       =       26.04
                                Prob > chi2      =       0.0005
Log likelihood = -25.794271      Pseudo R2    =       0.3355
```

low	Coef.	Std. Err.	z	P> z	[95% Conf. Interval]	
lwt	-.0183757	.0100806	-1.82	0.068	-.0381333	.0013819
smoke	1.400656	.6278396	2.23	0.026	.1701131	2.631199
ptd	1.808009	.7886502	2.29	0.022	.2622828	3.353735
ht	2.361152	1.086128	2.17	0.030	.2323796	4.489924
ui	1.401929	.6961585	2.01	0.044	.0374836	2.766375
race						
black	.5713643	.689645	0.83	0.407	-.7803149	1.923044
other	-.0253148	.6992044	-0.04	0.971	-1.39573	1.345101

We might prefer to see results presented as odds ratios. We could have specified the `or` option when we first fit the model, or we can now redisplay results and specify `or`:

```
. clogit, or
Conditional (fixed-effects) logistic regression

                                Number of obs   =       112
                                LR chi2(7)       =       26.04
                                Prob > chi2      =       0.0005
Log likelihood = -25.794271      Pseudo R2    =       0.3355
```

low	Odds Ratio	Std. Err.	z	P> z	[95% Conf. Interval]	
lwt	.9817921	.009897	-1.82	0.068	.9625847	1.001383
smoke	4.057862	2.547686	2.23	0.026	1.185439	13.89042
ptd	6.098293	4.80942	2.29	0.022	1.299894	28.60938
ht	10.60316	11.51639	2.17	0.030	1.261599	89.11467
ui	4.06303	2.828513	2.01	0.044	1.038195	15.90088
race						
black	1.770681	1.221141	0.83	0.407	.4582617	6.84175
other	.975003	.6817263	-0.04	0.971	.2476522	3.838573

Smoking, previous preterm delivery, hypertension, uterine irritability, and possibly the mother's weight all contribute to low birthweight. Race of black and race of other are statistically insignificant when compared with the race of white omitted group, although the race of black effect is large. We can test the joint statistical significance of race being black (2.race) and race being other (3.race) by using `test`:

```
. test 2.race 3.race
( 1) [low]2.race = 0
( 2) [low]3.race = 0

      chi2( 2) =    0.88
      Prob > chi2 = 0.6436
```



For a more complete description of `test`, see [R] `test`. `test` presents results in coefficients rather than odds ratios. Jointly testing that the coefficients on `2.race` and `3.race` are 0 is equivalent to jointly testing that the odds ratios are 1.

Here one case was matched to one control, that is, 1:1 matching. From `clogit`'s point of view, that was not important— $k_1$  cases could have been matched to  $k_2$  controls ( $k_1:k_2$  matching), and we would have fit the model in the same way. Furthermore, the matching can change from group to group, which we have denoted as  $k_{1i}:k_{2i}$  matching, where  $i$  denotes the group. `clogit` does not care. To fit the conditional logistic regression model, we specified the `group(varname)` option, `group(pairid)`. The case and control are stored in separate observations. `clogit` knew that they were linked (in the same group) because the related observations share the same value of `pairid`.

◀

## □ Technical note

`clogit` provides a way to extend McNemar's test to multiple controls per case (1: $k_{2i}$  matching) and to multiple controls matched with multiple cases ( $k_{1i}:k_{2i}$  matching).

In Stata, McNemar's test is calculated by the `mcc` command; see [R] `epitab`. The `mcc` command, however, requires that the matched case and control appear in one observation, so the data will need to be manipulated from 1 to 2 observations per stratum before using `clogit`. Alternatively, if you begin with `clogit`'s 2-observations-per-group organization, you will have to change it to 1 observation per group if you wish to use `mcc`. In either case, `reshape` provides an easy way to change the organization of the data. We will demonstrate its use below, but we direct you to [D] `reshape` for a more thorough discussion.

In [example 2](#), we used `clogit` to analyze the relationship between low birthweight and various characteristics of the mother. Assume that we now want to assess the relationship between low birthweight and smoking, ignoring the mother's other characteristics. Using `clogit`, we obtain the following results:

```
. clogit low smoke, group(pairid) or
Iteration 0:  log likelihood = -35.425931
Iteration 1:  log likelihood = -35.419283
Iteration 2:  log likelihood = -35.419282
Conditional (fixed-effects) logistic regression
                                                    Number of obs   =       112
                                                    LR chi2(1)      =         6.79
                                                    Prob > chi2     =       0.0091
Log likelihood = -35.419282                    Pseudo R2       =       0.0875
```

	low	Odds Ratio	Std. Err.	z	P> z	[95% Conf. Interval]
	smoke	2.75	1.135369	2.45	0.014	1.224347 6.176763

Let's compare our estimated odds ratio and 95% confidence interval with that produced by `mcc`. We begin by reshaping the data:

```
. keep low smoke pairid
. reshape wide smoke, i(pairid) j(low 0 1)
Data                long  ->  wide
-----
Number of obs.      112  ->    56
Number of variables   3   ->    3
j variable (2 values) low  -> (dropped)
xij variables:
                        smoke -> smoke0 smoke1
```

We now have the variables `smoke0` (formed from `smoke` and `low = 0`), recording 1 if the control mother smoked and 0 otherwise; and `smoke1` (formed from `smoke` and `low = 1`), recording 1 if the case mother smoked and 0 otherwise. We can now use `mcc`:

```
. mcc smoke1 smoke0
```

Cases	Controls		Total
	Exposed	Unexposed	
Exposed	8	22	30
Unexposed	8	18	26
Total	16	40	56

```
McNemar's chi2(1) =      6.53   Prob > chi2 = 0.0106
Exact McNemar significance probability = 0.0161
Proportion with factor
Cases      .5357143
Controls   .2857143   [95% Conf. Interval]
difference .25       .0519726   .4480274
ratio      1.875     1.148685   3.060565
rel. diff. .35       .1336258   .5663742
odds ratio 2.75     1.179154   7.143667   (exact)
```

Both methods estimated the same odds ratio, and the 95% confidence intervals are similar. `clogit` produced a confidence interval of [1.22, 6.18], whereas `mcc` produced a confidence interval of [1.18, 7.14].

□

## Use of weights

With `clogit`, weights apply to groups as a whole, not to individual observations. For example, if there is a group in your dataset with a frequency weight of 3, there are a total of three groups in your sample with the same values of the dependent and independent variables as this one group. Weights must have the same value for all observations belonging to the same group; otherwise, an error message will be displayed.

▷ Example 3

We use the example from the above discussion of the `mcc` command. Here we have a total of 56 matched case–control groups, each with one case matched to one control. We had 8 matched pairs in which both the case and the control are exposed, 22 pairs in which the case is exposed and the control is unexposed, 8 pairs in which the case is unexposed and the control is exposed, and 18 pairs in which they are both unexposed.

With weights, it is easy to enter these data into Stata and run `clogit`.

```
. clear
. input id case exposed weight
      id      case  exposed  weight
1.  1 1 1 8
2.  1 0 1 8
3.  2 1 1 22
4.  2 0 0 22
5.  3 1 0 8
6.  3 0 1 8
7.  4 1 0 18
8.  4 0 0 18
9.  end

. clogit case exposed [w=weight], group(id) or
(frequency weights assumed)
Iteration 0:  log likelihood = -35.425931
Iteration 1:  log likelihood = -35.419283
Iteration 2:  log likelihood = -35.419282
Conditional (fixed-effects) logistic regression

                                Number of obs   =       112
                                LR chi2(1)       =         6.79
                                Prob > chi2      =       0.0091
                                Pseudo R2       =       0.0875

Log likelihood = -35.419282
```

case	Odds Ratio	Std. Err.	z	P> z	[95% Conf. Interval]	
exposed	2.75	1.135369	2.45	0.014	1.224347	6.176763



**Fixed-effects logit**

The fixed-effects logit model can be written as

$$\Pr(y_{it} = 1 \mid \mathbf{x}_{it}) = F(\alpha_i + \mathbf{x}_{it}\beta)$$

where  $F$  is the cumulative logistic distribution

$$F(z) = \frac{\exp(z)}{1 + \exp(z)}$$

$i = 1, 2, \dots, n$  denotes the independent units (called “groups” by `clogit`), and  $t = 1, 2, \dots, T_i$  denotes the observations for the  $i$ th unit (group).

Fitting this model by using a full maximum-likelihood approach leads to difficulties, however. When  $T_i$  is fixed, the maximum likelihood estimates for  $\alpha_i$  and  $\beta$  are inconsistent (Andersen 1970; Chamberlain 1980). This difficulty can be circumvented by looking at the probability of  $\mathbf{y}_i = (y_{i1}, \dots, y_{iT_i})$  conditional on  $\sum_{t=1}^{T_i} y_{it}$ . This conditional probability does not involve the  $\alpha_i$ , so they are never estimated when the resulting conditional likelihood is used. See Hamerle and Ronning (1995) for a succinct and lucid development. See *Methods and formulas* for the estimation equation.

#### ► Example 4

We are studying unionization of women in the United States by using the union dataset; see [XT] xt. We fit the fixed-effects logit model:

```
. use http://www.stata-press.com/data/r15/union, clear
(NLS Women 14-24 in 1968)

. clogit union age grade not_smsa south black, group(idcode)
note: multiple positive outcomes within groups encountered.
note: 2,744 groups (14,165 obs) dropped because of all positive or
      all negative outcomes.
note: black omitted because of no within-group variance.

Iteration 0:  log likelihood = -4521.3385
Iteration 1:  log likelihood = -4516.1404
Iteration 2:  log likelihood = -4516.1385
Iteration 3:  log likelihood = -4516.1385

Conditional (fixed-effects) logistic regression

                                Number of obs   =    12,035
                                LR chi2(4)       =     68.09
                                Prob > chi2      =     0.0000
                                Pseudo R2        =     0.0075

Log likelihood = -4516.1385
```

union	Coef.	Std. Err.	z	P> z	[95% Conf. Interval]	
age	.0170301	.004146	4.11	0.000	.0089042	.0251561
grade	.0853572	.0418781	2.04	0.042	.0032777	.1674368
not_smsa	.0083678	.1127963	0.07	0.941	-.2127088	.2294445
south	-.748023	.1251752	-5.98	0.000	-.9933619	-.5026842
black	0 (omitted)					

We received three messages at the top of the output. The first one, “multiple positive outcomes within groups encountered”, we expected. Our data do indeed have multiple positive outcomes ( $\text{union} = 1$ ) in many groups. (Here a group consists of all the observations for a particular individual.)

The second message tells us that 2,744 groups were “dropped” by `clogit`. When either  $\text{union} = 0$  or  $\text{union} = 1$  for all observations for an individual, this individual’s contribution to the log-likelihood is zero. Although these are perfectly valid observations in every sense, they have no effect on the estimation, so they are not included in the total “Number of obs”. Hence, the reported “Number of obs” gives the effective sample size of the estimation. Here it is 12,035 observations—only 46% of the total 26,200.

We can easily check that there are indeed 2,744 groups with `union` either all 0 or all 1. We will generate a variable that contains the fraction of observations for each individual who has `union = 1`.

```
. by idcode, sort: generate fraction = sum(union)/sum(union < .)
. by idcode: replace fraction = . if _n < _N
(21,766 real changes made, 21,766 to missing)
. tabulate fraction
```

fraction	Freq.	Percent	Cum.
0	2,481	55.95	55.95
.0833333	30	0.68	56.63
.0909091	33	0.74	57.37
.1	53	1.20	58.57
<i>(output omitted)</i>			
.9	10	0.23	93.59
.9090909	11	0.25	93.84
.9166667	10	0.23	94.07
1	263	5.93	100.00
Total	4,434	100.00	

Because  $2481 + 263 = 2744$ , we confirm what `clogit` did.

The third warning message from `clogit` said “black omitted because of no within-group variance”. Obviously, `race` stays constant for an individual across time. Any such variables are collinear with the  $\alpha_i$  (that is, the fixed effects), and just as the  $\alpha_i$  drop out of the conditional likelihood, so do all variables that are unchanging within groups. Thus they cannot be estimated with the conditional fixed-effects model.

There are several other estimators implemented in Stata that we could use with these data:

```
cloglog ... , vce(cluster idcode)
logit ... , vce(cluster idcode)
probit ... , vce(cluster idcode)
scobit ... , vce(cluster idcode)
xtcloglog ...
xtgee ... , family(binomial) link(logit) corr(exchangeable)
xtlogit ...
xtprobit ...
```

See [R] [cloglog](#), [R] [logit](#), [R] [probit](#), [R] [scobit](#), [XT] [xtcloglog](#), [XT] [xtgee](#), [XT] [xtlogit](#), and [XT] [xtprobit](#) for details.

## Stored results

`clogit` stores the following in `e()`:

### Scalars

<code>e(N)</code>	number of observations
<code>e(N_drop)</code>	number of observations dropped because of all positive or all negative outcomes
<code>e(N_group_drop)</code>	number of groups dropped because of all positive or all negative outcomes
<code>e(k)</code>	number of parameters
<code>e(k_eq)</code>	number of equations in <code>e(b)</code>
<code>e(k_eq_model)</code>	number of equations in overall model test
<code>e(k_dv)</code>	number of dependent variables
<code>e(df_m)</code>	model degrees of freedom
<code>e(r2_p)</code>	pseudo- <i>R</i> -squared
<code>e(ll)</code>	log likelihood
<code>e(ll_0)</code>	log likelihood, constant-only model
<code>e(N_clust)</code>	number of clusters
<code>e(chi2)</code>	$\chi^2$
<code>e(p)</code>	significance
<code>e(rank)</code>	rank of <code>e(V)</code>
<code>e(ic)</code>	number of iterations
<code>e(rc)</code>	return code
<code>e(converged)</code>	1 if converged, 0 otherwise

### Macros

<code>e(cmd)</code>	<code>clogit</code>
<code>e(cmdline)</code>	command as typed
<code>e(depvar)</code>	name of dependent variable
<code>e(group)</code>	name of <code>group()</code> variable
<code>e(multiple)</code>	multiple if multiple positive outcomes within group
<code>e(wtype)</code>	weight type
<code>e(wexp)</code>	weight expression
<code>e(title)</code>	title in estimation output
<code>e(clustvar)</code>	name of cluster variable
<code>e(offset)</code>	linear offset variable
<code>e(chi2type)</code>	Wald or LR; type of model $\chi^2$ test
<code>e(vce)</code>	<i>vce</i> type specified in <code>vce()</code>
<code>e(vcetype)</code>	title used to label Std. Err.
<code>e(opt)</code>	type of optimization
<code>e(which)</code>	max or min; whether optimizer is to perform maximization or minimization
<code>e(ml_method)</code>	type of ml method
<code>e(user)</code>	name of likelihood-evaluator program
<code>e(technique)</code>	maximization technique
<code>e(properties)</code>	<code>b V</code>
<code>e(predict)</code>	program used to implement <code>predict</code>
<code>e(marginsok)</code>	predictions allowed by <code>margins</code>
<code>e(marginsnotok)</code>	predictions disallowed by <code>margins</code>
<code>e(marginsdefault)</code>	default <code>predict()</code> specification for <code>margins</code>
<code>e(asbalanced)</code>	factor variables <code>fvset</code> as <code>asbalanced</code>
<code>e(asobserved)</code>	factor variables <code>fvset</code> as <code>asobserved</code>

### Matrices

<code>e(b)</code>	coefficient vector
<code>e(Cns)</code>	constraints matrix
<code>e(ilog)</code>	iteration log (up to 20 iterations)
<code>e(gradient)</code>	gradient vector
<code>e(V)</code>	variance-covariance matrix of the estimators
<code>e(V_modelbased)</code>	model-based variance

### Functions

<code>e(sample)</code>	marks estimation sample
------------------------	-------------------------

## Methods and formulas

Breslow and Day (1980, 247–279), Collett (2003, 251–267), and Hosmer, Lemeshow, and Sturdivant (2013, 243–268) provide a biostatistical point of view on conditional logistic regression. Hamerle and Ronning (1995) give a succinct and lucid review of fixed-effects logit; Chamberlain (1980) is a standard reference for this model. Greene (2018, chap. 18) provides a straightforward textbook description of conditional logistic regression from an economist’s point of view, as well as a brief description of choice models.

Let  $i = 1, 2, \dots, n$  denote the groups and let  $t = 1, 2, \dots, T_i$  denote the observations for the  $i$ th group. Let  $y_{it}$  be the dependent variable taking on values 0 or 1. Let  $\mathbf{y}_i = (y_{i1}, \dots, y_{iT_i})$  be the outcomes for the  $i$ th group as a whole. Let  $\mathbf{x}_{it}$  be a row vector of covariates. Let

$$k_{1i} = \sum_{t=1}^{T_i} y_{it}$$

be the observed number of ones for the dependent variable in the  $i$ th group. Biostatisticians would say that there are  $k_{1i}$  cases matched to  $k_{2i} = T_i - k_{1i}$  controls in the  $i$ th group.

We consider the probability of a possible value of  $\mathbf{y}_i$  conditional on  $\sum_{t=1}^{T_i} y_{it} = k_{1i}$  (Hamerle and Ronning 1995, eq. 8.33; Hosmer, Lemeshow, and Sturdivant 2013, eq. 7.4),

$$\Pr(\mathbf{y}_i \mid \sum_{t=1}^{T_i} y_{it} = k_{1i}) = \frac{\exp(\sum_{t=1}^{T_i} y_{it} \mathbf{x}_{it} \beta)}{\sum_{\mathbf{d}_i \in S_i} \exp(\sum_{t=1}^{T_i} d_{it} \mathbf{x}_{it} \beta)}$$

where  $d_{it}$  is equal to 0 or 1 with  $\sum_{t=1}^{T_i} d_{it} = k_{1i}$ , and  $S_i$  is the set of all possible combinations of  $k_{1i}$  ones and  $k_{2i}$  zeros. Clearly, there are  $\binom{T_i}{k_{1i}}$  such combinations, but we need not count all of these combinations to compute the denominator of the above equation. It can be computed recursively.

Denote the denominator by

$$f_i(T_i, k_{1i}) = \sum_{\mathbf{d}_i \in S_i} \exp\left(\sum_{t=1}^{T_i} d_{it} \mathbf{x}_{it} \beta\right)$$

Consider, computationally, how  $f_i$  changes as we go from a total of 1 observation in the group to 2 observations to 3, etc. Doing this, we derive the recursive formula

$$f_i(T, k) = f_i(T - 1, k) + f_i(T - 1, k - 1) \exp(\mathbf{x}_{iT} \beta)$$

where we define  $f_i(T, k) = 0$  if  $T < k$  and  $f_i(T, 0) = 1$ .

The conditional log-likelihood is

$$\ln L = \sum_{i=1}^n \left\{ \sum_{t=1}^{T_i} y_{it} \mathbf{x}_{it} \beta - \log f_i(T_i, k_{1i}) \right\}$$

The derivatives of the conditional log-likelihood can also be computed recursively by taking derivatives of the recursive formula for  $f_i$ .

Computation time is roughly proportional to

$$p^2 \sum_{i=1}^n T_i \min(k_{1i}, k_{2i})$$

where  $p$  is the number of independent variables in the model. If  $\min(k_{1i}, k_{2i})$  is small, computation time is not an issue. But if it is large—say, 100 or more—patience may be required.

If  $T_i$  is large for all groups, the bias of the unconditional fixed-effects estimator is not a concern, and we can confidently use `logit` with an indicator variable for each group (provided, of course, that the number of groups does not exceed `matsize`; see [R] [matsize](#)).

This command supports the clustered version of the Huber/White/sandwich estimator of the variance using `vce(robust)` and `vce(cluster clustvar)`. See [P] [\\_robust](#), particularly *Maximum likelihood estimators* and *Methods and formulas*. Specifying `vce(robust)` is equivalent to specifying `vce(cluster groupvar)`, where *groupvar* is the variable for the matched groups.

`cllogit` also supports estimation with survey data. For details on VCEs with survey data, see [SVY] [variance estimation](#).

## References

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## Also see

- [R] [cllogit postestimation](#) — Postestimation tools for `cllogit`
- [R] [asclogit](#) — Alternative-specific conditional logit (McFadden’s choice) model
- [R] [logistic](#) — Logistic regression, reporting odds ratios
- [R] [mlogit](#) — Multinomial (polytomous) logistic regression
- [R] [nlogit](#) — Nested logit regression
- [R] [ologit](#) — Ordered logistic regression
- [R] [scobit](#) — Skewed logistic regression
- [BAYES] [bayes: cllogit](#) — Bayesian conditional logistic regression
- [MI] [estimation](#) — Estimation commands for use with `mi` estimate
- [SVY] [svy estimation](#) — Estimation commands for survey data
- [XT] [xtgee](#) — Fit population-averaged panel-data models by using GEE
- [XT] [xtlogit](#) — Fixed-effects, random-effects, and population-averaged logit models
- [U] [20 Estimation and postestimation commands](#)